

THE COMPLETE ASSIGNMENT OF THE  $^{13}\text{C}$  NMR SPECTRA  
OF LASALOCID AND THE SODIUM SALT-COMPLEX  
OF THE ANTIBIOTIC

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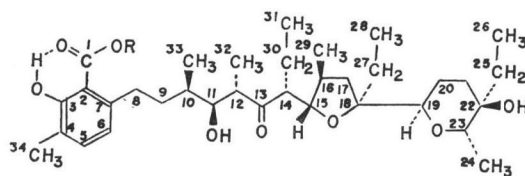
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All thirty-four signals observed in the  $^{13}\text{C}$  nmr of both the free acid form (**Ia**) and sodium salt (**Ib**) of the polyether antibiotic lasalocid have been assigned. This was achieved using model compounds such as 3-methylsalicylic acid, the retroaldol ketones from both lasalocid and lysocellin and a  $\gamma$ -lactone from a third polyether, salinomycin. The last assignments to be made were accomplished using biosynthetically enriched samples of the antibiotic.

One of the earliest biosynthetic applications of  $^{13}\text{C}$  nmr spectroscopy in the field of natural products was a study<sup>1)</sup> that led to the first *bona fide* demonstration of butyric acid incorporation, by *Streptomyces lasaliensis* in the formation of the three C-ethyl groups in lasalocid (**Ia**). This discovery was later reported for another polyether antibiotic, monensin A<sup>2)</sup>. A more recent utility for the  $^{13}\text{C}$  nmr technique has been found in the study of the solution conformation of complex organic compounds such as salinomycin<sup>3)</sup> and lysocellin<sup>4)</sup>, which also belong to the polyether class of antibiotics.

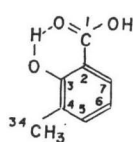
In this report, the partial assignments of lasalocid's  $^{13}\text{C}$  nmr spectrum made from earlier biosynthetic studies<sup>1,5)</sup> are completed. The results are presented for both the free acid, ethanolate (**Ia**) and the sodium salt-complex (**Ib**) and provide preliminary data for the eventual conformational analysis of lasalocid and its salts in solution by  $^{13}\text{C}$  nmr. Conformational analyses of **Ia** have been

Fig. 1. Numbering system for compounds **I**, **II**, **III**, **IV** and **V** based on the lasalocid nomenclature (ref. 10).

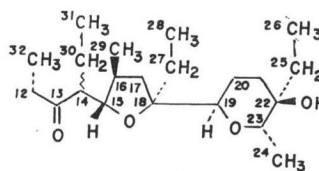


**Ia:** R = H,  $\text{C}_2\text{H}_5\text{OH}$ ; Lasalocid A, ethanolate.

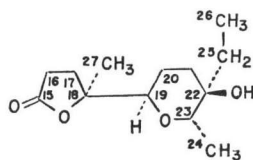
**Ib:** R = Na; Lasalocid A, sodium salt-complex.



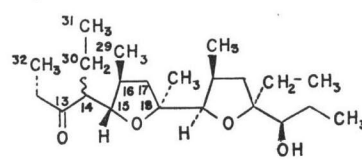
**III**



**II**



**IV**



**V**

reported using proton nmr spectroscopy<sup>6,7)</sup> and our results should provide information to complement these analyses. For instance, a cursory examination of the results presented in Table 1 reveals that three of the seven quaternary carbons, C-1, 2 and 13 (see Fig. 1) are the three which undergo the largest downfield shift on sodium complexation.

#### Assignment of the <sup>13</sup>C NMR of the Lasalocid Retroaldol Ketone, II

Pyrolysis<sup>8)</sup> or base treatment<sup>9)</sup> of **Ia** (or **Ib**) produces *inter alia* the ketone **II**, a C<sub>21</sub>H<sub>34</sub>O<sub>4</sub> molecule in which the C-14 center<sup>10)</sup> is epimerized but the other six asymmetric centers are unaffected. This result was expected on chemical grounds and was confirmed by <sup>13</sup>C nmr which gives double peaks for ten of the twelve carbons adjacent to C-14 whereas the eight carbons associated with the terminal tetrahydro-pyran all give single peaks. The paired signals are assigned by partially relaxed FOURIER transform (PRFT) spectra<sup>11)</sup> assuming the relaxation times of similar carbons in both epimers are roughly equal (Table 1).

Table 1. Assignment of the <sup>13</sup>C nmr spectra of lasalocid free acid **Ia** and sodium salt complex **Ib** together with the retroaldol ketone **II**

Carbon number <sup>a</sup>	Functional group	<sup>13</sup> C Shift in ppm <sup>b</sup>		
		<b>Ia</b> (in CD <sub>2</sub> Cl <sub>2</sub> )	<b>Ib</b> (in CD <sub>2</sub> Cl <sub>2</sub> )	<b>II</b> (in CDCl <sub>3</sub> )
1	-CO <sub>2</sub> H	173.6	176.4	
2	=C-	111.2	118.2	
3	=C-OH	161.6	161.3	
4	=C-CH <sub>3</sub>	124.1	123.0	
5	=CH	135.1	131.5	
6	=CH	121.6	119.6	
7	=C-CH <sub>2</sub>	144.4	143.5	
8	-CH <sub>2</sub>	34.8	33.4	
9	-CH <sub>2</sub>	37.0	38.1	
10	-CH(CH <sub>3</sub> )	34.9	34.5	
11	-CH(OH)	72.8	71.0	
12	-CH(CH <sub>3</sub> )	48.9	49.0	37.1 (36.4)
13	C=O	214.1	219.9	212.6
14	-CH(C <sub>2</sub> H <sub>5</sub> )	55.4	55.9	58.9 (57.2)
15	-CH-O	84.2	83.1	86.5 (85.7)
16	-CH(CH <sub>3</sub> )	34.8	34.1	38.3 (36.9)
17	-CH <sub>2</sub>	38.7	37.9	41.0 (40.7)
18	-C(O)	86.7	87.6	84.4 (84.3)
19	-CH(O)	70.9	68.6	72.9
20	-CH <sub>2</sub>	20.0	19.5	21.3
21	-CH <sub>2</sub>	30.2	29.2	29.5
22	-C(O)	72.4	71.5	70.5
23	-CH(O)	76.6	77.2	76.6
24	-CH <sub>3</sub>	14.0	13.6	14.2
25	-CH <sub>2</sub>	30.7	31.1	30.6
26	-CH <sub>3</sub>	6.6	6.7	6.4
27	-CH <sub>2</sub>	30.6	29.8	28.7 (28.4)
28	-CH <sub>3</sub>	9.2	9.5	8.1
29	-CH <sub>3</sub>	15.9	16.0	17.6 (16.7)
30	-CH <sub>2</sub>	16.7	16.2	21.3 (21.0)
31	-CH <sub>3</sub>	12.9	12.4	12.5 (12.2)
32	-CH <sub>3</sub>	13.2	12.6	7.4
33	-CH <sub>3</sub>	13.4	13.3	
34	-CH <sub>3</sub>	15.7	15.3	

<sup>a</sup> For the numbering system, see Fig. 1.

<sup>b</sup> Downfield from internal Me<sub>4</sub>Si.

The resonance of 212.6 ppm is characteristic of a ketone carbon (C-13) and comparison of **II** with the  $\gamma$ -lactone, **IV** (Table 2) obtained from salinomycin<sup>3)</sup> facilitates the assignment of the eight carbons, C-19 to C-26 in **II** as shown in Table 1. By elimination, the remaining oxymethine (C-15) and quaternary carbon (C-18) are assigned to the peaks at 86.7 (85.7)\* and 84.6 (84.3) ppm respectively. The carbons, C-12 and C-14, adjacent to the carbonyl can be assigned using deuterium exchange, and this leaves one methine peak at 38.3 (36.9) ppm, due to C-16. Carbons C-17, 29, 30, 31 and 32 of the lasalocid retroaldol ketone **II** can be conveniently assigned by comparison with the retroaldol ketone **V** (Table 2) from lysocellin solved earlier<sup>12)</sup>. The remaining methylene carbon at  $\delta$ 28.7 (28.4) ppm and the methyl at 8.1 ppm arise from the ethyl C-27 and C-28 respectively. Due to the proximity of C-29 and C-31 to the epimerized center at C-14, both peaks due to these methyls are doubled, whereas the four other methyls in **II**; C-24, 26, 28 and 32 give single peaks as indicated in Table 1.

#### Assignment of <sup>13</sup>C Spectrum of Lasalocid A, Ethanolate

Crystallization of lasalocid A free acid from ethanol yields a monoethanolate, **Ia** (C<sub>34</sub>H<sub>53</sub>O<sub>8</sub>·

Table 2. Assignment of the <sup>13</sup>C nmr spectra of model compounds related to lasalocid

Carbon number <sup>a</sup>	Functional group	$\delta$ - <sup>13</sup> C Shift in ppm <sup>b</sup> of <b>III</b> , <b>IV</b> & <b>V</b> in CDCl <sub>3</sub>		
		<b>III</b>	<b>IV</b>	<b>V</b>
1	-CO <sub>2</sub> H	173.4		
2	=C-	112.6		
3	=C-OH	160.8		
4	=C-CH <sub>3</sub>	126.7		
5	=CH	136.8		
6	=CH	118.9		
7	=C-CH <sub>2</sub>	[128.6] <sup>c</sup>		
12	-CH(CH <sub>3</sub> )			37.3 (37.1)
13	C=O			213.6
14	-CH(C <sub>2</sub> H <sub>5</sub> )			59.3 (57.2)
15	-CH(O)		[177.3] <sup>c</sup>	87.3 (86.7)
16	-CH(CH <sub>3</sub> )		29.6	36.9 (36.4)
17	-CH <sub>2</sub>		29.1	42.5
18	-C(O)		87.3	
19	-CH(O)		73.0	
20	-CH <sub>2</sub>		21.2	
21	-CH <sub>2</sub>		29.0	
22	-C(O)		70.8	
23	-CH(O)		76.9	
24	-CH <sub>3</sub>		14.3	
25	-CH <sub>2</sub>		30.7	
26	-CH <sub>3</sub>		6.3	
27	-CH <sub>2</sub>		[22.9] <sup>c</sup>	
28	-CH <sub>3</sub>			
29	-CH <sub>3</sub>			16.8 (16.2)
30	-CH <sub>2</sub>			21.5 (21.0)
31	-CH <sub>3</sub>			12.6 (12.2)
32	-CH <sub>3</sub>			7.3
34	-CH <sub>3</sub>	15.5		

<sup>a</sup> For the carbon numbering system, see Fig. 1.

<sup>b</sup> Downfield from internal Me<sub>4</sub>Si.

<sup>c</sup> Functionalities of these carbons are different from those of the corresponding carbons in **Ia**, **Ib** and **V**.

\* Where two peaks were observed for a single carbon, the one appearing at higher field is arbitrarily expressed in parentheses. Therefore, chemical shift values in parentheses do not necessarily correspond to one epimer.

C<sub>2</sub>H<sub>6</sub>O). The assignment of the <sup>13</sup>C nmr spectrum of **Ia** was accomplished by comparison with the retroaldol ketone, **II** (*vide supra*), and 2-hydroxy-3-methyl-benzoic acid (**III**), which was assigned by selective proton decoupling. Both of these models are assigned as shown in Table 2. Single frequency off-resonance decoupling experiments were employed in the assignment of all the carbons between C-13 to C-31 in **Ia**, except C-17 and C-19. Although the chemical shift of these two carbons in **II** suggested the peaks at δ 38.7 and 72.8 ppm as arising from C-17, 19 in **Ia**, biosynthetic enrichment<sup>5)</sup> results (Table 3) showed that although the signal at δ 38.7 was due to C-17, the correct assignment of the C-19 peak is at δ 70.9 ppm. Of the remaining seven carbons; C-8 to C-12, C-32 and C-33, the peak at δ 48.9 ppm is assigned to C-12\* due to the characteristic chemical shift for a methine adjacent to a ketone. The assignment of C-9 and C-11 (like C-19) was deduced from the biosynthetic enrichments summarized in Table 3. This implies that the remaining methine peak at δ 34.9 corresponds to C-10 and the remaining methylene peak at δ 34.8 ppm corresponds to C-8. The last of the methyl peaks at δ 13.2 and 13.4 ppm are tentatively assigned to C-32 and C-33 respectively, completing the assignment of **Ia**. From these results, together with our earlier proposals<sup>5)</sup> for **Ib**, the complete assignment of the latter can be achieved. It is noteworthy that on going from **Ia** to **Ib**, the order of shifts is the same except for carbons C-9, 17 and C-11, 22 (Table 1).

Examining the change in chemical shifts (Table 1) for the free acid (**Ia**) on conversion to the sodium salt (**Ib**), the most obvious downfield shifts are observed for carbons C-2 (7.0 ppm), C-13 (5.8 ppm) and C-1 (2.8 ppm) whereas upfield shifts on sodium complexation are greatest for C-5 (3.6 ppm), C-19 (2.3 ppm) and C-6 (2.0 ppm). Results using other cation complexes are needed before any meaningful conclusions can be drawn regarding the solution conformations of **Ia** and **Ib** by <sup>13</sup>C nmr spectroscopy.

Table 3. Incorporation results from **Ia** used to assign the <sup>13</sup>C nmr peaks at δ 37.0, 38.7, 70.9 and 72.8 ppm

Carbon number in <b>Ia</b>	Incorporation of [1- <sup>13</sup> C] acetate, [1- <sup>13</sup> C] propionate and [1- <sup>13</sup> C] butyrate into lasalocid as determined by <sup>13</sup> C nmr of the free acid			
	<sup>13</sup> C Shift in ppm in CD <sub>2</sub> Cl <sub>2</sub>	% Abundance* of <sup>13</sup> C in lasalocid produced from		
		CH <sub>3</sub> <sup>13</sup> CO <sub>2</sub> Na	CH <sub>3</sub> CH <sub>2</sub> <sup>13</sup> CO <sub>2</sub> Na	CH <sub>3</sub> CH <sub>2</sub> CH <sub>2</sub> <sup>13</sup> CO <sub>2</sub> Na
9	37.0	1.0	<u>3.5</u>	1.0
17	38.7	1.0	1.0	<u>4.0</u>
19	70.9	<u>2.0</u>	1.0	1.0
11	72.8	1.0	<u>3.5</u>	1.0

\* Corrected to nearest 0.5%.

### Experimental

The <sup>13</sup>C nmr spectra of lasalocid A sodium salt<sup>1)</sup> and γ-lactone<sup>3)</sup> have been reported previously. The <sup>13</sup>C nmr spectrum of lasalocid A free acid ethanolate was run on a Varian XL-100 spectrometer (25.2 MHz) in CD<sub>2</sub>Cl<sub>2</sub> and those of the retroaldol ketones (in CDCl<sub>3</sub>) and 2-hydroxy-3-methyl-benzoic acid (in CD<sub>3</sub>OD) on a JEOL FX-100 spectrometer (25.05 MHz).

Deuteration of **II** was carried out under the same conditions as for the retroaldol reaction of **I**<sup>5)</sup> with H<sub>2</sub>O being replaced by D<sub>2</sub>O.

\* In an earlier paper (ref. 5), this signal was assigned to C-8. Single frequency off-resonance decoupling experiments, however, showed the signal was due to a methine and therefore the assignment is revised as shown in Table 1.

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